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Biopolymer-Technologie Aktiengesellschaft

Protocol

Specification

Prepito DNA Blood250 Kit

for general purposes

DNA purification from up to 250 µl blood

Kit Components

Magnetic Beads
Lysis Buffer
Binding Buffer
Wash Buffer
Wash Buffer
Wash Buffer
Wash Buffer
Elution Buffer
Protease

Deep Well Plates
Single Tubes
Disposable Tips

Completion time: approximately 55 minutes

Typical yield: 5 - 10 µg DNA

Storage Conditions and Safety Information

This kit may be stored at room temperature (15 – 25 °C) and is stable for at least 1 year following delivery. The kit buffers contain irritant substances. Take appropriate laboratory safety measures and wear gloves when handling.

The included protocol is sufficient for most blood samples: fresh, non-coagulated and frozen. This kit is optimized for DNA purification from human blood samples obtained from healthy individuals.

Using this method 0.5 - 2 % of the eluate is normally a sufficient template for PCR amplification.

it's chemagic!

Version 071127

Any further questions?

chemagen technical support **Fon** +49 (0) 2401 805-50 **Mail** support@chemagen.com

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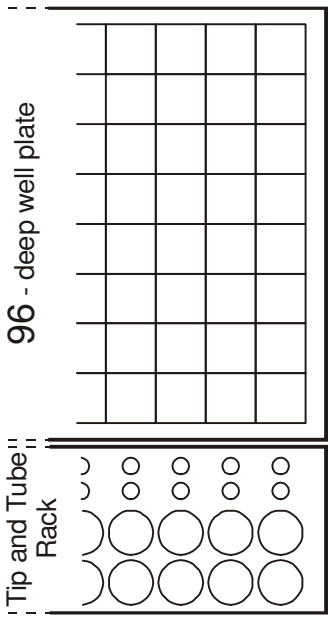
Positioning Procedure

see "Protocol Steps"for detailed information



100 - 250 µl whole blood and **Protease***

Pos. 4 second row for Disposable Tips; **! not used in this protocol !**
Pos. 3 Disposable Tips

Pos. 2 single tubes with 150 µl **Magnetic Beads**

Pos. 1 single tubes, empty (elution)

! **** Protease is required only for sample volumes > 150 µl whole blood***

Before You Start

1. Dissolve **Protease** in the appropriate volume of distilled water (see protease flask label).
2. Connect all buffer supply containers to the **chemagic Prepito**. Take care that all buffer supply containers contain enough buffer for the selected number of samples (see and follow the instructions in the manual).

! ***The reconstituted Protease is stable for 2 months at 2 – 8 °C or at –20 °C.***

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Protocol Steps

1. Press [**change protocol**].
2. Select the [**Blood 250**] protocol.
3. Press [**continue**].
4. Enter your 4-digit access code for authorization and confirm by pressing [**enter**].
5. Confirm that the correct protocol is chosen by pressing [**enter**].
6. Confirm the protocol description by pressing [**continue**].
7. Choose the positions where the samples will be placed and confirm by pressing [**continue**].
8. Enter the kit barcode and confirm by pressing [**ok**].
9. If sample tracking is required press [**yes**] and follow the instructions on the touch screen panel to enter the accordant barcodes. If no sample tracking is required press [**no**].
10. Place all plastic materials to the appropriate positions. Place one empty tube (position 1), one tube filled with 150 µl of **Magnetic Beads** (position 2) and one disposable tip (position 3) for each sample to positions according to the sample positions.
11. If using blood volumes more than 250 µl add 10 µl of **Protease** to each well of the Deep Well Plate (DWP, riplate SW) defined as sample well (see section above “Positioning Procedure”). For sample volumes less or equal than 150 µl do not add any **Protease**.
12. Add up to 250 µl of the blood sample to each sample well prefilled with **Protease**.

! ***Incubation of the blood/Protease mixtures longer than 5 minutes can lead to lower yields and decreased purities of the extracted DNA. Therefore continue immediately with further protocol steps after adding the blood samples.***

13. Ensure again that your samples are placed in the correct positions. Place the DWP on the tracking system and press [**continue**].
14. Place the **chemagic Tip & Tube Rack** to the correct position on the tracking system. Check for accurate fitting of the DWP and lock by closing the safety latch.
15. Close the cover and immediately start the automated isolation process by pressing [**start**].

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General Remarks

The **Elution Buffer** included in this kit is 10 mM Tris-HCl pH 8.0. TE buffer pH 8.0 can also be used without any protocol adjustments. Water pH 8.0 may also be used, but the yield could be slightly decreased.

The **Magnetic Bead** suspension should be mixed vigorously before dispensing, otherwise the suspension is not homogenous and the DNA yield could be low.

UV Measurements

In some cases you may find traces of magnetic beads left in the eluate. Such particles will not interfere with PCR and most downstream applications but may increase the background UV measurements. In such a case, prior to UV analysis, we recommend an additional separation step using a manual separator in order to separate any traces of particles.

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